

Short communication

Brief Communication: Caffeic Acid Derivatives and Polymethoxylated Flavonoids from Cat's Whiskers (*Orthosiphon stamineus*) Form Stable Complexes with SARS-CoV Molecular Targets: An *In silico* Analysis

Siti Nur Athirah Mohd Kaspi^{1,2}, Nisha Govender^{1*} and Zeti-Azura Mohamed-Hussein^{1,2}

¹Institute of Systems Biology (INBIOSIS), Universiti Kebangsaan Malaysia, 43600 UKM, Bangi, Selangor, Malaysia

²Faculty of Science and Technology, Universiti Kebangsaan Malaysia, 43600 UKM, Bangi, Selangor, Malaysia

ABSTRACT

Cat's whiskers or the 'misai kucing' is an herbal plant native to the Southeast Asian region. The polyphenol enriched leaf extract contains numerous medicinal properties of major pharmaceutical interest. In this study, selected cat's whiskers polyphenols were screened computationally to predict the minimum binding affinities with severe acute respiratory syndrome coronavirus (SARS-CoV) molecular targets. Molecular docking analysis showed that the caffeic acid derivatives and polymethoxylated flavonoids from cat's whiskers bound stably to the binding pocket regions of SARS-CoV molecular targets at -4.2 to -7.1 kcal/mol. Furthermore, these cat's whiskers polyphenol-bound SARS-CoV complexes were held fairly strongly by hydrophobic interactions, hydrogen bonds, and electrostatic interactions at various extents.

Keywords: CoV-2, COVID-19, misai kucing, molecular docking, molecular target, polyphenol, SARS- therapeutics

ARTICLE INFO

Article history:

Received: 29 September 2021

Accepted: 06 December 2021

Published: 24 January 2022

DOI: <https://doi.org/10.47836/pjtas.45.1.13>

E-mail addresses:

a164782@siswa.ukm.edu.my (Siti Nur Athirah Mohd Kaspi)

nishag@ukm.edu.my (Nisha Govender)

zeti.hussein@ukm.edu.my (Zeti-Azura Mohamed-Hussein)

* Corresponding author

INTRODUCTION

The coronavirus disease (COVID-19) first emerged in Wuhan, China, in late 2019. It has been more than a year since the pandemic emerged, and the disease continues to impact the socio-economic lives of the world population. At this time of writing (9th November 2021), more than 250 million people have been infected with COVID-19, and up to 5 million deaths

have been reported globally. Malaysia has reported 2.5 million COVID-19 cases and 29,291 deaths (www.worldometers.info).

COVID-19 is divided into three distinct phases, and each phase represents a unique clinical stage (Gu & Korteweg, 2007). The asymptomatic stage 1 (about 1-2 days upon infection) begins with the inhalation of airborne droplets of SARS-CoV-2. The virus binds to receptors localized at the epithelial cells in the nasal cavity. Stage 2 (2-5 days upon viral entry) marks the virus propagation at the upper airway and conducting airway, followed by migration heading downwards to the respiratory tract. During this stage, a robust innate immune response is triggered. About 80% of the COVID-19 infected patients experience mild symptoms when the virus manifestation becomes restricted to the upper and conducting airways (Wu & McGoogan, 2020). In stage 3, massive viral propagation develops into severe disease symptoms as a result of aggravated immune response (cytokine storm) (Tang et al., 2020).

Orthosiphon stamineus Benth. (*Clerodendrathus spicatus* Thunb.) is well-known as 'misai kucing' in Malaysia, 'java tea' in Indonesia and 'sen cha' in China (Abdullah et al., 2012; Li et al., 2021). It is one of the most widely grown and used medicinal plants in Asia, southern China, and Australia (Abdullah et al., 2012). The long-term consumption of *O. stamineus* extract (OE) has been shown to offer protection against various diseases (Pandey & Rizvi, 2009). For thousands of years, the leaves, in their dried and/or fresh form

are consumed as a tea to treat multiple ailments such as urinary tract infection (Deipenbrock & Hensel, 2019), arthritis (Tabana et al., 2016), diabetes mellitus, hypertension, tonsillitis, rheumatism, and menstrual disorders (Cai et al., 2020; Zhong et al., 2012). Bioactive compounds pre-dominantly found in the leaves of *O. stamineus* have been reported to display anti-allergic, anti-angiogenic, anti-hypertensive, anti-inflammatory, antioxidant, and diuretic properties (Ashraf et al., 2018, 2020; Muhammad et al., 2011; Tabana et al. 2016). Despite being an important folk medicine, the potential activities of the *O. stamineus* bioactive compounds for COVID-19 management are least reported. Generally, the *O. stamineus* leaves constitute a broad range of bioactive compounds: pentacyclic triterpenes, phenolics, and small sterols. The *O. stamineus* leaves are especially enriched in polyphenols: up to 0.5 to 0.7% of the leaf extract contains flavonoids with methoxylated flavones and prenylated flavones, among the majority fraction (Deipenbrock & Hensel, 2019). In Malaysia, *O. stamineus* leaf extract has been reported to contain sinensetin, eupatorine, rosmarinic acid, caffeic acid, and gallic acid (Engku-Hasmah et al., 2013; Muhammad et al., 2011). In others, chloroform extracts of *O. stamineus* leaves showed high contents of sinensetin and eupatorine flavonoids, whilst the ethanolic leaf extract was characterized with high rosmarinic acid (Tabana et al. 2016).

In this study, the caffeic acid derivatives and polymethoxylated flavonoids from *O.*

stamineus were docked with SARS-CoV molecular targets at the binding pocket regions, and the minimum binding affinities of the resulting complexes were evaluated. Structural proteins governing key biological functions of the different stages of the SARS-CoV life cycle were identified from Protein Data Bank (PDB). The ligand-protein complexes with good minimum binding affinities were identified and further visualized. Our findings highlight the potentials of *O. stamineus* polyphenols for COVID-19 management strategies such as phytotherapy, preventive therapeutics, and immune health-enhancing and illness relief beverages/supplements.

METHODS

Data Collection and Target Pre-Processing

In this study, four different SARS-CoV structural proteins were selected and retrieved from the PDB (www.rcsb.org): membrane protein (PDB ID: 3I6G), main protease (PDB ID: 5RE4), and spike glycoprotein (PDB ID: 6VXX) (PDB ID: 6VYB). The binding pocket regions expressed as x, y, and z coordinates in a 3D space were identified using Biovia Discovery Studio 2021 version 4.5. In its native format (.PDB), each protein was manually pre-processed using the AutoDock tools described by Trott and Olson (2010); crystal water molecules were discarded, and hydrogen polar atoms and Kollman charges were added. The edited files were stored in the protein data bank, partial charge (Q), and atom type (T) (PDBQT) format.

The 3D structures of the following *O. stamineus* bioactive compounds (hereafter, designated as ligands) were searched and retrieved from PubChem (pubchem.ncbi.nlm.nih.gov): sinensetin (PubChem Compound ID number [CID]: 145659), gallic acid (PubChem CID: 370), caffeic acid (PubChem CID: 689043), rosmarinic acid (PubChem CID: 5281792), and eupatorine (PubChem CID: 97214). First, the structure data (SDF) format files were converted to PDB format using Open Babel Compiler version 2.3.2. Next, the ligand files were pre-processed using the AutoDock tools (Trott & Olson, 2010): set the number of torsions and detect the root atoms. Following the edits, all files were saved in the PDBQT format.

Ligand-Receptor Docking

A configuration file defining the binding pocket regions (grid box) of the receptors was prepared in txt. file. The selected ligands (sinensetin, gallic acid, caffeic acid, rosmarinic acid, and eupatorine) were docked against the binding pocket regions of the receptors using AutoDock Vina 1.1.2. (Trott & Olson, 2010). All docking runs were initiated at Command Prompt (Windows operating system). The output log files were evaluated, and the best-docked conformations were selected based on binding energy (kcal/mol), cluster RMSD value, number of stabilizing interactions such as hydrogen bonds, and other weak interactions. The 3D complex structure and interactions were visualized using the Biovia Discovery Studio version 4.5.

RESULTS AND DISCUSSION

The coronavirus disease-2019 (COVID-19) is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). The SARS-CoV-2 pathogenesis mechanism is governed by structural proteins (surface spike protein, membrane protein, envelope protein, and main protease-replicase polyproteins) and a few non-structural proteins (nsp6, nsp8, and nsp10) (Walls et al., 2020). The global outbreak is one of the most challenging pandemics ever experienced, and effective drugs are yet to avail (November 2021). However, numerous research efforts, especially promising and high-throughput computational biology approaches for plant-based therapeutics, have emerged through molecular docking, modeling, and dynamic simulation studies (Skariyachan et al., 2020).

The *O. stamineus* is a traditional medicinal herbal plant with rich bioactive compounds. Extensively concentrated in the leaves, there are a large number of different types of bioactive compounds identified in *O. stamineus* (Li et al., 2021; Tezuka et al., 2000). As such, the total phenolic content of *O. stamineus* leaf 50% ethanol extract was 385 µg/ml, and the total flavonoid content, 57 µg/ml (Tabana et al., 2016). Rosmarinic acid is the main constituent of *O. stamineus* leaves. Previous studies have reported that *O. stamineus* leaves contain two different polyphenols in abundance: polymethoxylated flavonoids and caffeic acid derivatives (Engku-Hasmah et al., 2013; Muhammad et al., 2011; Tabana et al., 2016). This study selected the most

abundantly occurring polyphenols in the *O. stamineus* leaves as ligands. Sinestine and eupatorine are polymethoxylated flavonoids, while rosmarinic acid, gallic acid, and caffeic acid belong to the caffeic acid derivatives group (Olah et al., 2003).

Four different structural SARS-CoV-related proteins were selected, namely the membrane protein (PDB ID: 3I6G), main proteases (PDB ID: 5RE4), and spike glycoproteins (PDB ID: 6VXX and 6VYB). The biological roles of the selected proteins are described as follows: 3I6G is a SARS-CoV M protein complexed with HLA-A*0201 obtained from X-ray diffraction method; 5RE4 is a crystal structure of SARS-CoV-2 main protease complexed with Z1129283193 obtained by X-ray diffraction method; 6VXX is a closed state model of SARS-CoV-2 spike glycoprotein obtained via electron microscopy, and 6VYB, an open state of SARS-CoV-2 spike ectodomain structure obtained through electron microscopy (www.rcsb.org).

The minimum binding affinities for all possible combinations of ligand (sinensetin, gallic acid, caffeic acid, rosmarinic acid, and eupatorine)-protein complexes were measured at root mean square deviation (RSMD)=0. For complexes formed between the ligands and the 3I6G, the binding affinity ranged from -5.3 to -7.1 kcal/mol (Table 1). A various number of hydrogen bonds, hydrophobic, and electrostatic interactions were observed at the receptor residue (RR) of all 3I6G-bound complexes except the 3I6G-gallic acid complex (absence of

hydrophobic interaction) (Table 2). The rosmarinic acid-3I6G complex showed the highest binding affinity at -7.1 kcal/mol, followed by sinensetin-3I6G complex at -6.6 kcal/mol (Table 1). In contrast, the number of interactions was higher in the sinensetin-3I6G complex as compared to the rosmarinic acid-3I6G complex. There were three hydrogen bonds (RR: ARG97, TYR123, HIS114), three hydrophobic interactions (RR: VAL152, TRP147, ALA150), and one electrostatic interaction (ASP77) in the rosmarinic acid-3I6G complex. In comparison, the sinensetin-3I6G complex showed three hydrogen bonds (RR: TYR7, GLU63, LYS66), six hydrophobic interactions (RR: TYR7, TYR159, PHE9, HIS70, TRP167, LYS66), and a single electrostatic interaction (RR: LYS66) (Figure 1A) (Table 2). On the other hand, 3I6G-gallic acid showed the lowest binding affinity at 5.3 kcal/mol with a single hydrogen bond (RR: TRP14) and electrostatic interaction (RR: ASP77) (Tables 1 and 2).

The minimum binding affinities of ligands bound to 5RE4 ranged from -4.9 to -6.7 kcal/mol. The rosmarinic acid-5RE4

complex shows the least minimum binding affinity at -6.7 kcal/mol, followed by the eupatorine-5RE4 and sinensetin-5RE4 complexes at -5.7 kcal/mol and -5.6 kcal/mol, respectively (Table 1). Although the rosmarinic acid-5RE4 complex showed no presence of electrostatic interactions, a total of nine hydrogen bonds (RR: ARG76, LEU75, PHE66, ASP92, THR93, HIS64, LEU67, LEU75, VAL77) along two hydrophobic interactions (RR: ARG76, VAL73) were observed (Figure 1B). The eupatorine-5RE4 complex showed the presence of a hydrogen bond (RR: VAL77), hydrophobic interaction (RR: ARG76), and electrostatic (RR: ARG76) interaction each. Besides the rosmarinic acid-5RE4 complex, both the gallic acid-5RE4 and caffeic acid-5RE4 complexes showed an absence of electrostatic interaction (Table 2).

For the ligand-6VXX complexes, the minimum binding affinities range at -4.3 to -5.6 kcal/mol (Table 1), and electrostatic interactions were consistently absent in all complexes (Table 2). The eupatorine-6VXX complex showed the least minimum binding affinity at -5.6 kcal/mol with three hydrogen bonds (RR: PHE86, ASN87, ASN234)

Table 1
Molecular docking analysis of Orthosiphon stamineus polyphenols and SARS-CoV molecular targets (proteins) performed using the Autodock Vina software

	3I6G	5RE4	6VXX	6VYB
Sinensetin	-6.6	-5.6	-4.9	-4.8
Gallic acid	-5.3	-4.9	-4.7	-4.5
Caffeic acid	-5.4	-5.0	-4.3	-4.2
Rosmarinic acid	-7.1	-6.7	-5.5	-6.0
Eupatorine	-6.5	-5.7	-5.6	-5.6

Note. The first row denotes receptor ID (Protein Data Bank), and the first column represents polyphenols. All numerical values indicate the minimum binding affinities expressed in kcal/mol

Table 2

The interactions between the Orthosiphon stamineus polyphenol and receptor residue

Receptor-ligand complex	Hydrogen bond	Hydrophobic interaction	Electrostatic interaction
3I6G-sinensetin	TYR7, GLU63, LYS66	TYR7, TYR159, PHE9, HIS70, TRP167, LYS66	LYS66
3I6G-gallic acid	TRP14	-	ASP77
3I6G-caffeic acid	LYS66, ARG97, HIS114	TYR15	LYS66
3I6G-rosmarinic	ARG97, TYR123, HIS114	VAL152, TRP147, ALA150	ASP77
3I6G-eupatorine	TYR7, LYS66	TYR7, TYR159, LEU156, PHE9, LYS66	LYS66
5RE4-sinensetin	PHE66, LEU75	VAL73, LEU75, LEU67, ARG76	ARG76, ASP92
5RE4-gallic acid	GLN74, PHE66, VAL77	ARG76, VAL77	-
5RE4-caffeic acid	GLN74, PHE66, HIS64, VAL77	ARG76, VAL77	-
5RE4-rosmarinic	ARG76, LEU75, PHE66, ASP92, THR93, HIS64, LEU67, LEU75, VAL77	ARG76, VAL73	-
5RE4-eupatorine	VAL77,	ARG76	ARG76
6VXX-sinensetin	THR236, ILE233	ARG237, THR236	-
6VXX-gallic acid	ASN196, ILE237, ASN234, ILE235	-	-
6VXX-caffeic acid	-	-	-
6VXX-rosmarinic	ASP88, ASN196, THR236	-	-
6VXX-eupatorine	PHE86, ASN87, ASN234,	THR236, PHE86	-
6VYB-sinensetin	THR236, ASN234	THR236, ARG237	-
6VYB-gallic acid	THR108, LYS113, GLY232	ILE233	-
6VYB-caffeic acid	ILE235	-	-
6VYB-rosmarinic	THR236, ILE235	-	-
6VYB-eupatorine	-	-	-

and two hydrophobic interactions (RR: THR236, PHE86) (Figure 1C). The caffeic acid-6VXX complex showed the lowest binding affinity at -4.3 kcal/mol with no interactions (Table 1).

The minimum binding affinities for ligand-6VYB ranged from -4.2 to -6.0 kcal/mol. The rosmarinic acid-6VYB complex showed the least minimum binding affinity at -6.0 kcal/mol, followed by the eupatorine-6VYB complex at -5.6 kcal/mol. Caffeic acid-6VYB complex showed

the least minimum binding affinity (-4.2 kcal/mol) among all the ligand-6VYB complexes (Table 1). All the ligand-6VYB complexes had no electrostatic interaction. Interestingly, the sinensetin-6VYB complex showed a relatively lower minimum binding affinity (-4.8 kcal/mol) than the rosmarinic acid-6VYB complex (-6.0 kcal/mol) and the number of interactions displayed a *vice versa* trend. The rosmarinic acid-6VYB complex showed two hydrogen bonds (RR: THR236, ILE235) only, while the

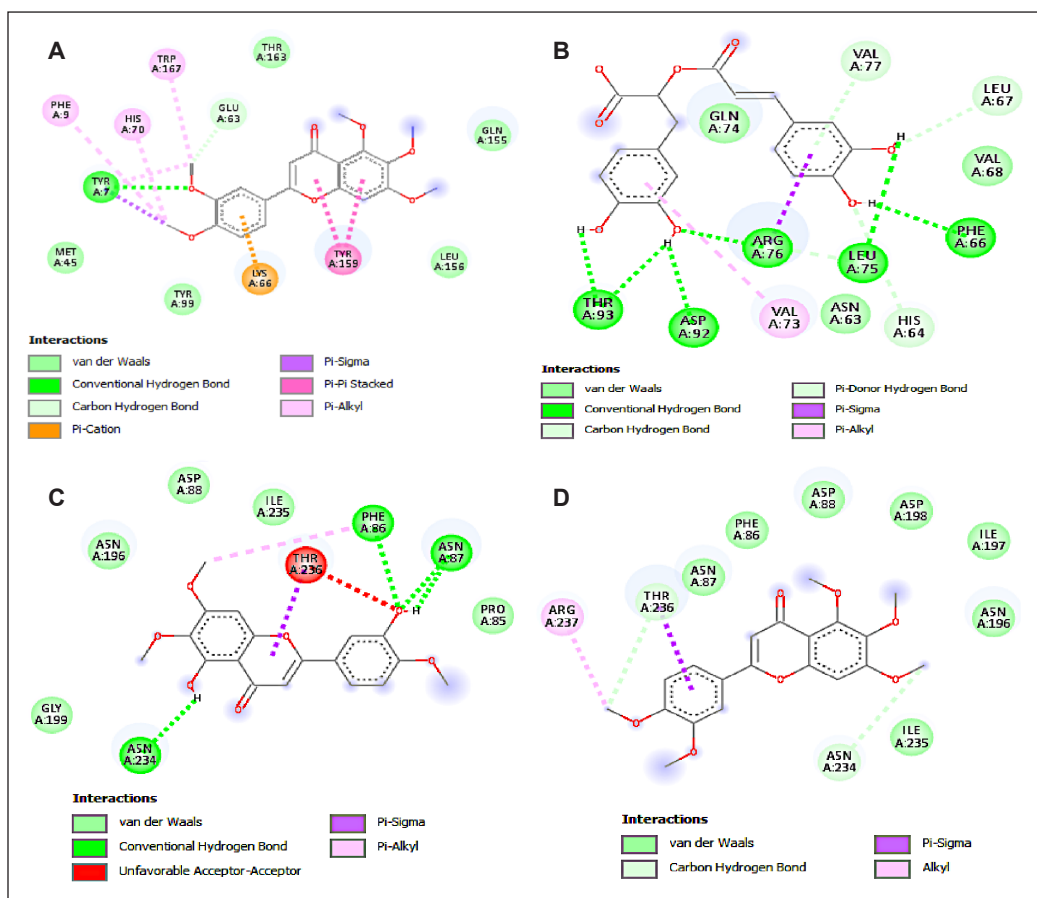


Figure 1. Interactions between the *Orthosiphon stamineus* polyphenols and SARS-CoV receptors. A) 316G-sinensetin complex; B) 5RE4-rosmarinic acid complex; C) 6VXX-eupatorine complex, and D) 6VYB-sinensetin complex. The best conformation complexes are selected based on the minimum binding affinity values and number of interactions at RMSD = 0. All 2- dimension *O. stamineus* polyphenol-bound SARS-CoV receptors complexes contain nodes, represent the receptor residue (RR), and dotted lines represent the interactions between ligand atom and RR

sinensetin-6VYB complex showed a greater number of interactions with two hydrogen bonds (RR: THR236, ASN234) and two hydrophobic interactions (RR: THR236, ARG237) (Figure 1D) (Table 2).

Plant-based natural products have been used to treat various diseases for a long time. These natural resources are mainly derived from beneficial traditional plant medicines. In time of the COVID-19

pandemic, the use of traditional medicinal plants as alternative medicine has been overwhelming, especially among Asian countries. Both China and the Indian government have recommended polyherbal decoctions for COVID-19 prevention and management: Qingfei Paidu, Huashi Baide, Xuanfei Badu, Toujie Quwen granules, Jinhua Qinggan granules (traditional Chinese medicine), Kabasura Kudineer

(traditional Siddha medicine). In Thailand, *Andrographis paniculata* (local name, ‘hempedu bumi’) is used for mild COVID-19 infection, whilst other herbal plants such as the lemon balm (*Melissa officinalis* L.), skullcap (*Scutellaria lateriflora*), Siberian ginseng (*Eleutherococcus senticosus*) are also widely employed throughout the world. As plant-based medicines pose minimal side effects compared to steroidal drugs, the ethnopharmacological relevance of traditional herbal plants for COVID-19 can be explored using extensive computational approaches, bioinformatics, and cheminformatics. In this study, five different polyphenols enriched in *O. stamineus* leaves (the sinensetin, caffeic acid, gallic acid, rosmarinic acid, and eupatorine) showed structural interactions with SARS-CoV proteins. In terms of enzymes encoding pathogenesis, the SARS-CoV is similar to SARS-CoV-2 (Lan et al., 2020). The structural insights of the potential interactions between the *O. stamineus* polyphenols and SARS-CoV proteins require further investigations to confirm and fully validate the inhibitory properties of *O. stamineus* polyphenols against SARS-CoV proteins. Other molecular dynamic simulation studies, drug-likeness analysis, and bioassay studies are necessary to identify the inhibitory and druggability roles of *O. stamineus* polyphenols against SARS-CoV-2 of COVID-19. The use of *O. stamineus* to treat multiple ailments could be stretched to viral diseases associated with SARS-CoV, such as the COVID-19.

CONCLUSION

The *in silico* analysis of *Orthosiphon stamineus* polyphenols against SARS-CoV molecular targets shows good interaction at a minimum binding affinity range of -4.2 to -7.1 kcal/mol coupled with numerous hydrogen bonds and hydrophobic interactions. The *O. stamineus* polyphenols are lucrative alternatives to synthetic drugs as they are comparably cheap and may pose fewer side effects in anti-viral treatments. With further research that complements the present preliminary findings, *O. stamineus* polyphenols could potentially be utilized for COVID-19 therapeutics development.

ACKNOWLEDGEMENTS

The first author would like to thank the Institute of Systems Biology (INBIOSIS), Universiti Kebangsaan Malaysia, Bangi, Selangor, Malaysia, for the internship training.

REFERENCES

- Abdullah, S., Shaari, A. R., & Azimi, A. (2012). Effect of drying methods on metabolites composition of misai kucing (*Orthosiphon stamineus*) leaves. *APCBEE Procedia*, 2, 178–182. <https://doi.org/10.1016/j.apcbee.2012.06.032>
- Ashraf, K., Halim, H., Lim, S. M., Ramasamy, K., & Sultan, S. (2020). *In vitro* antioxidant, antimicrobial and antiproliferative studies of four different extracts of *Orthosiphon stamineus*, *Gynura procumbens* and *Ficus deltoidei*. *Saudi Journal of Biological Sciences*, 21(1), 417-432. <https://doi.org/10.1016/j.sjbs.2019.11.003>
- Ashraf, K., Sultan, S., & Adam, A. (2018). *Orthosiphon stamineus* Benth. is an outstanding food medicine: Review of phytochemical and

- pharmacological activities. *Journal of Pharmacy and Bioallied Sciences*, 10(3), 109-118. https://doi.org/10.4103/jpbs.JPBS_253_17
- Cai, X., Zhang, L., Chen, X., Zhang, H., Xue, H., Lu, Y., Tang, J., & Lu, Y. (2020). *Orthosiphon stamineus* and rosmarinic acid reduce heat stress in laying hens. *Livestock Science*, 240, 104124. <https://doi.org/10.1016/j.livsci.2020.104124>
- Deipenbrock, M., & Hensel, A. (2019). Polymethoxylated flavones from *Orthosiphon stamineus* leaves as antiadhesive compounds against uropathogenic *E. coli*. *Fitoterapia*, 139, 104387. <https://doi.org/10.1016/j.fitote.2019.104387>
- Engku-Hasmah, E. A., Ahmad Tarmizi, S., Noor Ismawaty, N., & Abdul Ghani, O. (2013). Total phenolic, total flavonoid and rosmarinic acid contents in *Orthosiphon stamineus* (Misai Kucing) in relation to storage conditions. *Acta Horticulturae*, 1012, 837–842. <https://doi.org/10.17660/ActaHortic.2013.1012.113>
- Gu, J., & Korteweg, C. (2007). Pathology and pathogenesis of severe acute respiratory syndrome. *The American Journal of Pathology*, 170(4), 1136–1147. <https://doi.org/10.2353/ajpath.2007.061088>
- Lan, J., Ge, J., Yu, J., Shan, S., Zhou, H., Fan, S. (2020). *Crystal structure of the 2019-nCoV spike receptor-binding domain bound with ACE2 receptor*. <https://www.biorxiv.org/content/10.1101/2020.02.19.956235v1.full.pdf>
- Li, Z., Qu, B., Zhou, L., Chen, H., Wang, J., Zhang, W., & Chen, C. (2021). A new strategy to investigate the efficacy markers underlying the medicinal potentials of *Orthosiphon stamineus* Benth. *Frontiers in Pharmacology*, 12, 748684. <https://doi.org/10.3389/fphar.2021.748684>
- Muhammad, H., Gomes-Carneiro, M. R., Poa, K. S., De-Oliveira, A. C. A. X., Afzan, A., Sulaiman, S. A., Ismail, Z., & Paumgarten, F. J. R. (2011). Evaluation of the genotoxicity of *Orthosiphon stamineus* aqueous extract. *Journal of Ethnopharmacology*, 133(2), 647–653.
- Olah, N. K., Radu, L., Mogosan, C., Hanganu, D., & Gocan, S. (2003). Phytochemical and pharmacological studies on *Orthosiphon stamineus* Benth. (Lamiaceae) hydroalcoholic extracts. *Journal of Pharmaceutical and Biomedical Analysis*, 33(1), 117-123. [https://doi.org/10.1016/s0731-7085\(03\)00227-9](https://doi.org/10.1016/s0731-7085(03)00227-9)
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity*, 2, 897484. <https://doi.org/10.4161/oxim.2.5.9498>
- Skariyachan, S., Gopal, D., Chakrabarti, S., Kempanna, P., Uttarkar, A., Muddebihalkar, A. G., & Niranjana, V. (2020). Structural and molecular basis of the interaction mechanism of selected drugs towards multiple targets of SARS-CoV-2 by molecular docking and dynamic simulation studies—deciphering the scope of repurposed drugs. *Computational Biology Medicine*, 126, 104054. <https://doi.org/10.1016/j.compbimed.2020.104054>
- Tabana, Y. M., Al-Suede, F. S., Ahamed, M. B., Dahham, S. S., Hassan, L. E., Khalilpour, S., Taleb-Agha, M., Sandai, D., Majid, A. S. A., & Majid, A. M. S. A. (2016). Cat's whiskers (*Orthosiphon stamineus*) tea modulates arthritis pathogenesis via the angiogenesis and inflammatory cascade. *BMC Complementary and Alternative Medicine*, 16(1), 480. <https://doi.org/10.1186/s12906-016-1467-4>
- Tang, Y., Liu, J., Zhang, D., Xu, Z., Ji, J., & Wen, C. (2020). Cytokine storm in COVID-19: The current evidence and treatment strategies. *Frontiers in Immunology*, 11, 1708. <https://doi.org/10.3389/fimmu.2020.01708>
- Tezuka, Y., Stampoulis, P., Banskota, A. H., Awale, S., Tran, K. Q., Saiki, I., & Kadota, S. (2000). Constituents of the Vietnamese medicinal

- plant *Orthosiphon stamineus*. *Chemical and Pharmaceutical Bulletin*, 48(110),1711-1719.
- Trott, O., & Olson, A. J. (2010). AutoDock Vina: Improving the speed and accuracy of docking with new scoring function, efficient optimization and multithreading. *Journal of Computational Chemistry*, 31(2), 455-461. <https://doi.org/10.1002/jcc.21334>
- Walls, C., Park., Y., Tortorici, M. A., Wall, A., McGuire, A. T., & Veesler, D. (2020). Structure, function and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell*, 181(2), 281-292. <https://doi.org/10.1016/j.cell.2020.02.058>
- Wu, Z., & McGoogan, J. M. (2020). Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in China: Summary of a report of 72314 cases from the Chinese Center for Disease Control and Prevention. *JAMA - Journal of the American Medical Association*, 323(13), 1239–1242. <https://doi.org/10.1001/jama.2020.2648>
- Zhong, Y. S., Yu, C. H., Ying, H. Z., Wang, Z. Y., & Cai, H. F. (2012). Prophylactic effects of *Orthosiphon stamineus* Benth. extracts on experimental induction of calcium oxalate nephrolithiasis in rats. *Journal of Ethnopharmacology*, 144(3), 761-767.